



Sieć Badawcza Łukasiewicz - Institute of Biopolymers & Chemical Fibres

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Microbiological Laboratory

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Laboratory has put into effect the quality system in agree with PN-EN ISO/IEC 17025:2018-02

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Lodz, date: 10/06/2020

Copy No: 1

## Report of microbiological tests No 34/2020

ISO 22196:2011

Measurement of antibacterial activity on plastics and other non-porous surfaces.



Hotes:

1. The commissions results refer only to the examinated sample

2. Yest report belongs to the orderer and can be used only upon his agreement.

3. Without agreement of our laboratory this test report can be reprinted only as a whole.

4. The owner of the test report when attitues the results is obliged to inform that the results were obtained in Microbiological Laboratory of Siel Badawcza Lukasiawcz-Institute of Biopolymers and Ohenkal Pibras in Add which is accredited by PCA.

5. The orderer has the right to complaint in the sequence 38 days from the date of receiving the test report.

6. Lest report comprises the results of properties not included in accreditation scope PCA.





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## continuation of the Test Report No 34/2020 of 10/06/2020

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OBJECTIVE:	Determination of antibacterial effect of film samples	
CUSTOMER:	Media Ikonos Sp. z o.o., ul. Gosławicka 2d, 45-446 Opole	
TEST REPORT REFERES TO:	Order of 28/05/2020	
OBJEDT OF TESTING:	Film	
SAMPLES MARKED BY CUSTOMER:	Film Profiflex Pro GPT FX100+ Antibacterial	
RECIVED SAMPLES:		
TESTING METHOD:	ISO 22196:2011 Measurement of antibacterial activity on plastics and other non-porous surfaces.	
SAMPLING:	The customer	
SAMPLE RECEIVED:	29/05/2020	
DATE OF TEST:	01-04/06/2020	
DATE OF TEST:		
PRINCIPLE:	A suspension of E. coli i. S. aureus with known cell density is applied to the test sample. After 24 hours, the bacteria are washed away and the number of cells that survived contact with the sample is determined. This number is compared to the number of bacteria on the control sample and on this basis the antibacterial activity of the test sample is calculated.	





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SAMPLES SIZE:	Samples prepared by the client. Three replicate test specimens were used for each bacterial strain. The test surface of each sample was 50x50mm. In addition, square of 0.07 mm thick sterile polyethylene film were prepared. 40 x 40 mm squares were prepared to cover the test samples and 50x50 mm squares as a control film without antibacterial effect.
STERILIZATION:	Samples were disinfected by wiping with 70% cthanol. The samples were then drie at room temperature under sterile conditions.
PREPARATION OF BACTERIAL SUSPENSION:	E. coli and S. aureus bacteria were reduced-seeded in Petri dishes with NB medium before the start of the test and incubated 20h at 35 ° C. Then the cultur was taken with a loop and inoculated with NB medium diluted 1/500 in distille water. In this way, a bacterial suspension was made to inoculate the sample (inoculum). The density of the suspension was determined densitometrically on the McFarland scale, the number of cells was read from the standard curve and dilute to obtain an inoculum with a density corresponding to the number of bacteria from 2.5x10 <sup>5</sup> cfu / ml to 10x10 <sup>5</sup> cfu / ml.
INOCULATION OF THE SAMPLES:	The prepared bacterial suspension was inoculated with test samples in a ratio of 0. ml per sample and covered with polyethylene film so that the suspensions were evenly distributed on the surfaces of the test samples.
INCUBATION CONDITIONS:	Samples were incubated 24h at 35°C in a humid atmosphere.

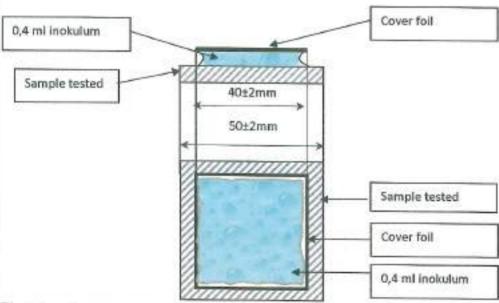


Fig. 1 Sample inoculation scheme according to ISO 22196: 2011





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#### EVALUATION OF BACTERIA GROWTH:

After incubation, the bacteria were washed out from the samples by shaking using 10 ml of SCDLP neutralizing fluid. Appropriate decimal dilutions were made from the obtained suspension. For each dilution, inoculation on agar plates (PCA medium) was performed and incubated 24h at 35°C. After incubation, colonies were counted on each plate and the number of bacteria was calculated using the formula:

$$N = (100 \times C \times V \times D) / A$$

where:

N - number of bacteria per cm2

C - arithmetic mean of all colonies from two plates from one dilution, where the plate must contain at least 30 colonies

D - dilution factor corresponding to the first calculated dilution (e.g. 102)

V - the number of milliliters of SDCLP used to wash the bacteria from the sample (10 ml)

A - surface of the cover foil in mm2

Antibacterial activity R for the test sample was calculated as:

$$R = (U_t - U_0) - (A_t - U_0) = U_t - A_t$$

where:

As - decimal logarithm of the average number of bacteria on the active sample (in triplicate) after 24 hours

U<sub>i</sub> - decimal logarithm of the average number of bacteria on the active control foil (in triplicate) after 24 hours

U<sub>0</sub> - decimal logarithm of the average number of bacteria on the active control foil (in triplicate) at time 0h.







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## continuation of the Test Report No 34/2020 of 10/06/2020 Test results for Escherichia coli ATCC 11229

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Date of test: 01-04/06/2020

Number of replicates: three replicates of each sample

Incubation temperature: 35°C

E. coli inoculum concentration: 7.9 x 105 cfu / ml.

Inoculum volume: 0.4ml

Conditions for a val	id test	
Requirements	Obtained results	Assessment of the study
The logarithmic value of the number of bacteria recovered immediately after inoculation from the untreated test specimens shall satisfy the following requiment: (Lmax-Lmin) / (Lmean) <0.2 where:  Lmax, Lmin - decimal logarithm of the largest and smallest number of bacteria on the sample immediately after inoculation of bacteria on the sample immediately after inoculation of bacteria on the sample immediately after inoculation of bacteria	L min = 4.17 L max = 4.22 Lmean = 4.20 (4.22-4.17)/(4.20) = 0.01	Meets the criteria
The average number of bacteria determined immediately after inoculation on the control sample should be within the range of 6.2x10 <sup>3</sup> to 2.5x10 <sup>4</sup> cfu/cm <sup>2</sup>	1.6x10 <sup>4</sup> cfu/cm <sup>2</sup>	Meets the criteria
The number of bacteria on each control sample after 24 hours of incubation should be not less than 6.1x10 <sup>1</sup> cfu/cm <sup>2</sup>	7.9x10 <sup>5</sup> cfu/cm <sup>2</sup> 9.3x10 <sup>5</sup> cfu/cm <sup>2</sup> 9.0x10 <sup>5</sup> cfu/cm <sup>2</sup>	Meets the criteria

Table 1. Antibacterial activity results of film samples against E. coli - results are given as the average of three replicates.

	Time [h]	Escherichia coli ATCC 11229		
Sample symbol		Number of bacteria average value N <sub>mean</sub> [cfu/cm <sup>2</sup> ]	log N <sub>mean</sub>	Antibacterial activity R
control	0	1.6 x 10 <sup>4</sup>	U <sub>0</sub> = 4.20	
control	24	8.8 x 10 <sup>5</sup>	Ut= 5.94	2
Film Profiflex Pro GPT FX100+ Antibacterial	24	5.5 x 10 <sup>3</sup>	A <sub>t</sub> = 3.72	2.22







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#### continuation of the Test Report No 34/2020 of 10/06/2020

Test results for Staphylococcus aureus ATCC 6538

Date of test: 01-04/06/2020

Number of replicates: three replicates of each sample

Incubation temperature: 35°C

S. aureus inoculum concentration: 4.2 x 105 cfu / ml,

Inoculum volume: 0.4ml

Conditions for a valid	l test	
Requirements	Obtained results	Assessment of the study
The logarithmic value of the number of bacteria recovered immediately after inoculation from the untreated test specimens shall satisfy the following requiment: (Lmax-Lmin) / (Lmax) <0.2 where:  Lmax, Lmin - decimal logarithm of the largest and smallest number of bacteria on the sample immediately after inoculation of bacteria on the sample immediately after inoculation of bacteria on the sample immediately after inoculation of bacteria on the	L min=4.08 L max=4.12 L <sub>mean</sub> =4.10 (4.12-4.08)/(4.10)= <b>0.01</b>	Meets the criteria
The average number of bacteria determined immediately after inoculation on the control sample should be within the range of 6.2x10 <sup>3</sup> to 2.5x10 <sup>4</sup> cfu/cm <sup>2</sup>	1.3x10 <sup>4</sup> efu/cm <sup>2</sup>	Meets the criteria
The number of bacteria on each control sample after 24 hours of incubation should be not less than 6.1x10 <sup>1</sup> cfu/cm <sup>2</sup>	5.2x10 <sup>4</sup> cfu/cm <sup>2</sup> 3.8x10 <sup>4</sup> cfu/cm <sup>2</sup> 4.5x10 <sup>4</sup> cfu/cm <sup>2</sup>	Meets the criteria

Table 2. Antibacterial activity results of film samples agains S. aureus - results are given as the average of three replicates.

	Time [h]	Staphylococcus aureus ATCC 6538		
Sample symbol		Number of bacteria average value N <sub>mean</sub> [cfu/cm <sup>2</sup> ]	log N <sub>mean</sub>	Antibacterial activity R
control foil	0	1.3 x 10 <sup>4</sup>	U <sub>0</sub> =4.10	+
control foil	24	4.5 x 10 <sup>4</sup>	U <sub>t</sub> = 4.65	2
Film Profiflex Pro GPT FX100+ Antibacterial	24	1.3 x 10 <sup>0</sup>	A <sub>4</sub> = 0.10	4.55

#### Conclusions:

The tested sample labeled Film Profiflex Pro GPT FX100+ Antibacterial showed antibacterial activity against tested strains of Escherichia coli ATCC 11229 and strong antibacterial activity with reference to Staphylococcus aureus ATCC 6538.

After 24 hours of incubation the number of E. coli bacteria in the control sample was 8.8 x 10<sup>5</sup> cfu/cm<sup>2</sup>, while on the test sample was 5.5 x 10<sup>5</sup> cfu/cm<sup>2</sup>. Antibacterial activity R was 2.22 with reference to E. coli.

After 24 hours of incubation the number of S. aureus bacteria in the control sample was 4.5 x 10<sup>4</sup> cfu/cm<sup>2</sup>, while on the test sample it was 1,3x10<sup>9</sup> cfu/cm<sup>2</sup>. Antibacterial activity R was 4.55 with reference to S. aureus.

The sample has antibacterial effect.

THE TEST PERFORMER: M. Sc. Eng Krystyna Guzińska

The date of report: 10/06/2020
The report drawn up in two identical copies:

copy 1 - the client receives copy 2 - remains at the contractor's AUTHORIZED: Head of Laboratory PhD Dorota Ka/mierczak

The end of the report